Journal of Plant Science & Research



Volume 6, Issue 2 - 2019 © Anu A, et al. 2019 www.opensciencepublications.com

Antitumor Activities of KALI Banana Fruit in MG-63 Cell Line

Research Article

Anu A and Geethalashmi S*

Department of Biotechnology, Sree Narayana Guru College, Coimbatore, India

***Corresponding author:** Geethalasmi S, Associate professor, Department of Biotechnology, Sree Narayana Guru College, Coimbatore, India; E-mail: s.geethalakshmi@gmail.com

Copyright: © Anu A, et al. 2019. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article Information: Submission: 09/10/2019; Accepted: 11/11/2019; Published: 13/11/2019

Abstract

Banana natural products are broadly utilized for human consumption. It is the second real nourishment crop in India. There are around 365 varieties of bananas available throughout the world. It is utilized as conventional medication for bowel disorders. This study aimed at examining the anticancer activity of Kali variety of banana which is commonly consumed in southern part of India. Anticancer examination of the sample was performed for the ethanolic extract using MG-63 cell lines which showed a high degree of anticancer activity which was proved by the cytotoxic effect on MG-63 tumor cell lines. Phytochemicals present in the plant concentrate provoked cell apoptosis and smoother cell expansion to quickly partitioning malignancy cell lines. The kali variety shows increased level of anti cancer activity in both cell lines. So we can use this variety in oral medicine for cancer treatment.

Keywords: MG-63; Kali banana fruit; Bioactive compounds; Apoptotic index; Phenols; Flavanoids

Introduction

Malignancy is a strange cell development with the potential to spread to other parts of the body (WHO, 2018). According to ASCO information, men have a high occurrence rate of bone marrow cancer than women. Genetic disorders, hereditary, exposure to toxic chemicals like solvents, fuels, certain cleaning products, radiation cause bone marrow cancer in human. There are 1660 death (960 men and 700 women) from this disease in 2018. A huge number of patients show resistance to tamoxifen, the currently available high potent drug for treating bone marrow cancer and experience extreme reactions. Other treatment methods like chemotherapy, radiation and bone marrow transplantation result in serious side effects which complicate the current situation. So, medical research focuses on natural methods to cure cancer without additional side effects. Numerous fruits like guava, banana [1], papaya, apple, watermelon, litchi have been accounted to have demonstrated therapeutic properties in several studies [1-6].

Kali is a local name of kali vazha which is a traditional variety of banana in South India. It is a wild type of banana and it cultivated in hill area in Kerala. This variety is used as traditional medicine in Kerala. The fruit occur as a bunch weighing 6-8 kg. Presently, this variety is endangered because of climatic change and modernization of agriculture techniques. The kali fruit is a rich source of crude protein (6-7%), crude fat (9-12%) and total ash (7.6%). Natural bioactive compounds such as steroids, flavonoids, tannins, phenol, Saponins, alkaloids, glycosides are found in the mature fruit. Pieces of banana have for some time been utilized in customary medication in Asia and Africa for treating wounds, decrease pain, for cell rejuvenation, antimicrobial agent and many more [4,6-9].

This study has been aimed at analyzing the anticancer activity of Kali fruit with reduced risk [5,8-12]. The present investigation was directed to profile the bioactive compounds in kali pulp and to evaluate its ability of cancer prevention and analyze its capacity to hinder the expansion of human bone osteosarcoma cells using MG-63 cell line.

Materials and Methods

Kali fruit was collected from Attapady, Palakkad district, Kerala, India. The peel was removed from fruit, the pulp was collected, shade dried at room temperature for 40 days, powdered and stored at 4 °C until further use.

Preparation of plant extract

Ten gram of dry fruit powder was dissolved in 100 ml of hexane in a conical flask and kept for 24 hours in a shaker. The mixture was then filtered twice using Whatman No. 1 filter paper and stored at 4°C.

Qualitative analysis of secondary metabolites

GC-MS investigation of plant concentrates was performed using Shimadzu GC-MS hardware of model QP 2010S containing Rxi-5 ms combined silica narrow section of 30 m length, 0.25 mm distance across and film thickness of 0.25 μ m. This analysis was done to find the metabolites present in the extract.

Cell line maintenance

Human MG63 cell lines were obtained from National Center for Cell Sciences (NCCS), Pune, India. Dulbecco's Modified Eagle Media (DMEM) was utilized for maintaining the cell line, which was enhanced with 10% Fetal Bovine Serum (FBS). Penicillin (100 U/ml) and streptomycin (100 μ g/ml) were added to the medium to prevent bacterial contamination. The medium with cell lines was kept in a humidified environment with 5% CO, at 37 °C.

Cytotoxicity assay

The MG-63 cells were placed in 24 well plates (1 X 105 cells per well) and incubated in 5% CO₂ environment at 37 °C. Cells (1X105/ well) were placed in 24-well plate and incubated in 37 °C with 5% CO₂ condition. Once the cells reached confluence, the prepared concentrations of extract (25 - 300 $\mu\text{g/ml})$ were added and kept in incubator for 24 hours. Then the samples were removed from the well and washed with phosphate-buffered saline (pH 7.4) or DMEM without serum. 0.5% of 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-Tetrazolium Bromide (MTT) was added to each well (100 µl/well) and incubated for 4 hours. Then 1 ml of Dimethyl Sulfoxide (DMSO) was added in all the wells to dissolve the formazan crystals. Each sample was placed in the cuvette; using DMSO as the blank the absorbance value at the wavelength of 570 nm was noted using Ultra-Violet (UV) Spectrophotometer in triplicate. The observed values were tabulated and the concentration required for 50% inhibition (IC₅₀) was determined graphically. The percentage cell viability was calculated by determining the ratio between treated MG-63 cells and control multiplied by 100.

Result and Discussion

The hexane extract of the fruit was analyzed for secondary metabolites using GC-MS analysis and the chromatogram and the list of bioactive compounds present in the extract (Figure 1 and Table 1).

The GC-MS chromatogram showed an intense peak at RT 9.376, a second large peak at RT 13.246 and least peak at RT 19.422. The larger peak showed a mass by charge ratio of 155.10 (Figure 1). This

m/z ratio does not correlate with any of the prior revealed anticancer bioactive mixes as per the verification in anticancer database and thus has all the earmarks of being a novel one.

The anticancer properties of the pulp with its tumor necrosis factor activity were already reported by many researchers [4,12,13]. MG-63 cell line is a model system for bone cancer studies and apoptosis process in cell. As per literature cited, with a high concentration of bioactive compounds like phenols and flavonoids, high intense anticancer activity can be observed on MG-63 cells [14-18]. Many treatments are used currently in cancer treatment like chemotherapy which, at primary stage, gives good response but it creates resistance in the later stage of cancer. In the present study, different concentration of fruit extract showed various levels of inhibitory action on MG-63 cell lines. Increase in concentration of the extract from 25 μ g /ml to 300 μ g /ml showed a decrease in cell viability of MG-63 from 90.14 to 26.7 % (Figures 2 and 3). The IC50 value of the fruit extract was found to be 170 μ g/ml [19-21].

Conclusion

From the present study, it is clear that the hexane concentrate of kali fruit extracts has a strong cytotoxic activity against MG-63 bone cancer. The fruit have broad spectrum of bioactive compounds which is anticancer active, which is seen from GC-MS chromatogram. Further research is needed to distinguish the bioactive compounds in banana, their refinement and interpretation of their activity to utilize the fruit pulp in pharmaceutical industries.

Table 1: Bioactive Constituents of kali fruit extract.

Peak	Retention time	COMPOUND	M.W. (Daltons)	Area %	Molecular formula
1	7.917	4H-pyran-4-one,2,3-Dihydro- 3,5-Dihydroxy-6-Methyl	101.05	11.99	$C_6H_8O_4$
2	8.248	Ethyl caprylate	88.10	1.82	C ₁₀ H ₂₀ O ₂
3	9.252	Coumaran	120.10	1.91	C ₈ H ₈ O
4	9.376	5-Hydroxymethylfurfural	97.05	30.09	$C_6H_6O_3$
5	9.708	3-Methoxycatechol	140.05	7.25	C7H8O3
6	11.086	Ethyl caprate	88.05	0.72	C8H16O2
7	11.230	5H-Imidazole-4-Carboxylic acid,5-Amino,Ethyl Ester	155.10	0.98	$C_4H_5N_3O_{2,1} \\ C_6H_{13}N_3O_{2}$
8	13.246	TrimethyloInitromethane	57.00	21.32	$C_4H_7N_4O_{11}$
9	16.048	3-Deoxy-d-mannoic lactone	57.00	20.75	$C_{6}H_{10}O_{5}$
10	19.422	Ethyl Palmitate	88.05	0.68	C18H36O2
11	30.272	Glycerol.beta-palmitate	57.05	2.48	$C_{3}H_{8}O_{3}$ $C_{16}H_{32}O_{2}$



Figure 1: GC- MS Chromatogram of Kali hexane extract.

JOURNAL OF PLANT SCIENCE & RESEARCH



Figure 2: Cytotoxicity assay indicating IC50 value.



Acknowledgement

The authors are grateful for the cooperation of the SCIGEN Research Lab, Staff of Department of Biotechnology, Sree Narayana Guru College for necessary support. Technical support from Dr. S. Geethalakshmi, Associate Professor and Head, Department of Biotechnology, Sree Narayana Guru College is also acknowledged.

References

- Rahmat A, Rosil R, Zain WN, Endrini S, Sani HA (2002) Ant proliferative activity of pure lycopene compared to both extracted lycopenes and juices from watermelon (*Citrullus vulgaris*) and papaya (*Carica papaya*) on human breast and liver cancer cell line. J Med Sci 2: 55-58.
- Arora M, Kaur P (2013) Antimicrobial and antioxidant activity of orange pulp and peel. Int J Sci Res 2: 412-415.
- Li ZY, Wang Y, Shen WT, Zhou P (2012) Content determination of benzyl glucosinolate and anti-cancer activity of its hydrolysis product in *Carica papaya* L. Asian Pac J Trop Med 72: 382-385.
- Liu RH (2004) potential synergy of phytochemicals in cancer prevention: mechanism of action. J Nutr 134: 3479S-3485S.

- Suresh Kumar P, Shiva KN, Mayil vaganan M, Uma S (2018) Waste utilization and functional foods from banana. Indian Hortic 63: 43-46.
- Tsamo CVP, Herent M, Tomekpe K, Emaga TH, Quentin Leclercq J, et al. (2015) Phenolic profiling in the pulp and peel of nine plantain cultivars (*Musa* sp.). Food Chem 15: 197-204.
- Manosroi J, Dhumtanom P, Manosroi A (2006) Anti-proliferative activity of essential oil extracted from Thai medical plants on KB and P388 cell line. Cancer Lett 235: 114-120.
- Orhue PO, Momoh AR (2013) Antibacterial activities of different solvent extracts of Carica Papaya fruit parts on some gram positive and gramnegative organisms. Int J Herbs Pharmacol Res 2: 42-47.
- Prasad KN, Hao J, Liu T, Li J, Wei X (2009) Antioxidant and anticancer activities of high Pressure assisted extract of longan (*Dimocarpus longan* Lour.) fruit pericarp. Innov Food Sci Emerg 10: 413-419.
- Ciniglia C,Pinto G, Sansone C, Pollio A (2010) Acridine orange/Ethidium bromide double stining test : A simple *In-vitro* assay to detect apoptosis induced by phenolic compounds in plant cells. Allelopathy J 26: 301-308.
- Kumar KPS, Bhowmik D, Duraivel S, Umadevi M (2012) Traditional and medicinal uses of banana. J pharmacogn Phytochem 1: 51-63.
- Kumar PS, Durgadevi S, Saravanan A, Uma S (2019) Antioxidant potential and antitumor activities of *Nendran* Banana peels in Breast cancer cell line. Indian J Pharma Sci 81: 464-473.
- Ghaasemi K, Ghasemi Y, Ebrahimzadeh MA (2009) Antioxidant activity, phenol and flavanoids contents of 13 citrus species peels and tissues. Pak J Pharm Sci 22: 227-281.
- Burda S, Oleszek W, and Lee CY (1990) Phenolic compounds and their changes in apples during maturation and cold storage. J Agric Food Chem 38: 945-948.
- Gonzalez Gallego J, Garcia Mediavilla MV, Sanchez Campos S, Tunon MJ (2010) Fruit polyphenols, immunity and inflammation. Br J Nutr 104: 15-27.
- Goulas V, and Manganaris G (2012) Exploring the phytochemical content and the antioxidant potential of Citrus fruits grown in Cyprus. Food Chem 131: 39-47.
- Ragu S, Faye G, Iraqui I, Masurel-Heneman A, Kolodner RD, et al. (2009) Oxygen metabolism and reactive oxygen species cause chromosomal rearrangements and cell death. Micro Biotechnol 104: 9747-9752.
- Kim H, Moon JY, Kim H, Lee DS, Cho M, et al. (2010) Antioxidant and antiproliferative activities of mango (*Magnifera indica* L.) flesh and peel. Food Chem 121: 429-436.
- Senthirlaraja P, Kathireasan K (2015) *In vitro* cytotoxicity MTT assay in Vero, HepG2 and MCF-7 cell lines study of marine Yeast. J Appl Pharm Sci 55: 80-84.
- Shixin D, West BJ, C, Jarakae JC (2010) A quantitative comparison of phytochemical components in global noni fruits and their commercial products. Food Chem 1: 267-270.
- 21. Unnati S, Ripal S, Sanjeev A, Niyati A (2013) Novel anticancer agents from plant sources. Chin J Nat Med 11: 16-23.